Biological control of Cactaceae in South Africa


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This review is a summary of developments that have contributed to the success of several biological control programmes against invasive cactus species (Cactaceae) that have been worked on in South Africa over the last 12 years. Six potential biological control agents have been identified for the control of Pereskia aculeata Mill. and molecular studies have identified the origin of the South African P. aculeata population. Host-specificity testing is now required for the three most promising of these agents. The successful biological control programme against Opuntia stricta (Haw.) Haw. has resulted in a change in management strategies against this weed in the Kruger National Park and the control of O. stricta is now almost entirely reliant on biological control. Taxonomic problems associated with the identification of Cylindropuntia fulgida var. fulgida (Engelm.) F.M.Knuth var. fulgida have been resolved and an appropriate cochineal insect (Hemiptera: Dactylopiidae) biotype has been released, resulting in substantial declines in Cyl. fulgida var. fulgida populations. A long-term monitoring programme has been initiated to evaluate the progress of this new cochineal insect biotype. The Harrisia mealybug, Hypoecoccus pungens Granara de Willink (Hemiptera: Pseudococcidae), which was originally released on Harrisia martinii (Labour.) Britton & Rose has been collected and redistributed onto Cereus jamacaru DC., where it reduces fruit production and leads to the death of both seedlings and large plants. Resolving taxonomic problems to ensure the correct identification of plant species and the most appropriate biological control biotypes have been key issues that have led to the successful control of several cactus weed species in South Africa.

Key words: cactaceous weeds, Pereskia aculeata, Opuntia stricta, Cylindropuntia fulgida, Cereus jamacaru, cochineal insects, biotypes.

INTRODUCTION

The first biological control agent deliberately introduced into South Africa against an invasive plant was the cochineal insect, Dactylopius ceylonicus (Green) (Hemiptera: Dactylopiidae), for the control of the weed, Opuntia monacantha Haw. (Cactaceae), in 1913 (Lounsbury 1915). The biological control programme was extremely successful, reducing the density of the weed to negligible levels (Zimmermann et al. 2009). Since this early success, a further 14 biological-control-agent entities (species or biotypes) have been released for the control of 15 cactaceous weed species in South Africa. The agents were originally released against eight target species (including O. monacantha) but subsequently became established on seven additional cactus species. Of the Cactaceae that have biological control agents established on them, eight species (53 %) are considered to be under substantial biological control (i.e. other forms of management are still needed to supplement biological control, but far less, or less frequently than before), and four species (27 %) are considered to be under complete biological control (i.e. no other management interventions are necessary) (Klein 2011). Notable successes include the biological control of Opuntia aurantiaca Lindl. (Moran & Annecke 1979; Moran & Zimmermann 1991a), Opuntia ficus-indica (L.) Mill. (Annecke & Moran 1978; Zimmermann & Moran 1991), and Opuntia stricta (Haw.) Haw. (Hoffmann et al. 1999), which are considered to be under substantial control; and Cylindropuntia leptocaulis (DC.) F.M.Knuth (Moran & Zimmermann 1991b), Cylindropuntia fulgida (Engelm.)
F.M.Knuth var. fulgida, Cereus jamacaru DC. (Klein 1999), and Harrisia martini (Labour.) Britton & Rose (Klein 1999), which are under complete control.

Factors that have contributed to the success of biological control of Cactaceae in South Africa include unusual aspects of the biology of the plants and of the cactophagous insects. The taxonomic isolation and characteristic morphology and anatomy of the Cactaceae have resulted in adaptations in their insect herbivores which in turn have resulted in almost no cactophagous insects being able to feed on any plant species outside of the family (Mann 1969; Moran 1980). Because there are no native Cactaceae in the Old World, with the possible exception of Rhipsalis baccifera (J.Müller) Stern, oligophagous cactophages can be used for biological control without risk to non-target native or commercial plants (but see below), thereby considerably broadening the pool of suitable agents that are available. The increasing commercial importance of some cactus species such as O. ficus-indica, Opuntia robusta Wendel and Hylocereus species (Cactaceae) in South Africa has hampered efforts to extend biological control of cactus weeds, especially for species in the genus Opuntia. This is not a new phenomenon. The introduction of Cactoblastis cactorum Berg. (Lepidoptera: Pyralidae: Phycitinae) into South Africa in 1933 was opposed initially on the grounds that the larvae would damage Opuntia plants which were providing fruit and fodder for commercial gain. It eventually took a parliamentary decision, in the early 1900s, to allow release of the moth on the grounds that the losses caused by O. ficus-indica far exceeded any gains that were derived from this and related cactus species (Annecke & Moran 1978; van Sittert 2002; Zimmermann et al. 2009). The success of C. cactorum in Australia helped sway opinion in favour of its release in South Africa (Pettey 1948; Annecke & Moran 1978; Beinart 2003).

The historical successes of many biological control programmes around the world have benefited from collaboration in the transfer of successful agents between countries sharing common problems. This approach has been especially prominent between Australia and South Africa in the case of the biological control of Cactaceae (Moran & Zimmermann 1984; Zimmermann et al. 2009). Not only have these exchanges assisted in the control of already widespread cactus species in South Africa (e.g. O. aurantiaca and O. ficus-indica), they have also facilitated the control of species that had not yet become problematic in South Africa by providing agents which had proved to be successful in Australia, e.g. H. martini (Hoffmann 1995).

In this paper, the biological control programmes against weeds in the family Cactaceae that have been worked on since 1999 in South Africa are reviewed, including the selection of potential biological control agents for Pereskia aculeata Mill., recent research on Cylindropuntia fulgida var. fulgida, long-term post-release evaluations on O. stricta, and some observations on the biological control of Cereus jamacaru. Species of Cactaceae that have not been worked on in the past 12 years are not included as they were dealt with in previous reviews (Annecke & Moran 1978; Moran & Annecke 1979; Moran & Zimmermann 1991a,b; Zimmermann & Moran 1991; Klein 1999).

**PERESKIA ACULEATA**

*Pereskia aculeata* (Barbados gooseberry, pereskia) is a primitive creeping cactus with well developed leaves and woody stems and branches (Fig. 1). The plant is armed with paired, hooked thorns at the leaf-bases on the young, green shoots and clusters of long spines which develop progressively on older, woody stems (Henderson 1995). Flowers are creamy-white and have creamy-white stamen filaments in South Africa, and either creamy-white or purple stamen filaments in the native distribution (Leuenberger 1986; Paterson et al. 2009). The fruits are yellow and spiny when immature, but as the fruit ripens the spines are lost and the fruit changes to an orange colour (Leuenberger 1986).

*Pereskia aculeata* is native to parts of Central America and the Caribbean as well as to southeastern Brazil and northern Argentina (Leuenberger 1986). It was first recorded in South Africa in the Cape Town Botanical Gardens in 1858 (McGibbon 1858) and has become naturalized in areas around Cape Town and in Gauteng, Mpumalanga and Limpopo provinces. It has also naturalized along the eastern coast of the country in the Eastern Cape Province and KwaZulu-Natal Province, where it is particularly invasive and abundant in forests and coastal vegetation (Fig. 2) (Henderson 2001). The plant overgrows and kills indigenous coastal and forest flora, sometimes even causing large trees to collapse under the weight of the...
infestation (Moran & Zimmerman 1991b, Paterson et al. 2011). It was declared a weed in 1979, Proclamation R35 (1979), and under the Conservation of Agricultural Resources Act (Act 43 of 1983) (CARA 1983). It is also listed as an invasive alien plant in the draft regulations in terms of the National Environmental Management: Biodiversity Act (Anon. 2004).

Mechanical and herbicidal control methods are not effective against *P. aculeata* and have deleterious effects on non-target plants because *P. aculeata* usually grows intertwined with native vegetation. Mechanical control is ineffective because any small piece of the plant that is overlooked can regenerate. Herbicides are not translocated within the plant’s tissues so all parts of the plant must be sprayed for effective control (Klein 1999). Biological control is therefore the only economically viable and sustainable method to control *P. aculeata* (Moran & Zimmermann 1991b).

The disjunct native distribution of *P. aculeata* was seen as a possible constraint to the biological control programme because insects or pathogens may have developed close associations with different *P. aculeata* populations (Paterson et al. 2009). This could result in some insects and pathogens being more damaging on plants from certain populations (Kniskern & Rausher 2001). *Pereskia aculeata* is

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**Fig. 1. Pereskia aculeata.** (Drawn by G. Condy; first published in Henderson (1995), ARC-Plant Protection Research Institute, Pretoria.)

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a polymorphic species that includes a number of different wild and garden varieties, and plants from the northern and southern regions within its native distribution are morphologically distinct (Leuenberger 1986). Plants from the South African populations exhibit a mixture of morphological characters from both regions of the native distribution making it difficult to determine the origin of the South African plants. A recent genetic study indicated that the South African *P. aculeata* population matched the population in Rio de Janeiro Province in southern Brazil most closely, making this region the most appropriate in which to commence surveys for biological control agents in terms of genotype matching (Paterson et al. 2009).

**Current biological control of *Pereskia aculeata***

The only biological control agent released for the control of *P. aculeata* is *Phenrica guerini* Bechyné (Coleoptera: Chrysomelidae) which was released in small numbers (from 36 to about 100 beetles per release) at ten sites in KwaZulu-Natal Province and at two sites in Eastern Cape Province between 1991 and 1997 (Klein 1999). Establishment was recorded at only two of the original release sites (30°24'S 33°41'E; 33°36'S 26°53'E) and no reduction in *P. aculeata* density was apparent at either site. Recent releases of higher numbers have resulted in the establishment of *P. guerini* at Wewe Dam (29°32'27"S 31°08'06"E) in KwaZulu-Natal Province and the high levels of damage at this site suggest that the potential impact of *P. guerini* may have been underestimated (D. Conlong, pers. comm.).

**Surveys for new potential biological control agents**

*Pereskia aculeata* has been surveyed for potential biological control agents in southern Brazil, northern Argentina, Venezuela and Dominican Republic during eight field trips between 1984 and 2007. Many of these field trips were brief and in some cases *P. aculeata* was only examined on an opportunistic basis while surveying for insects on various other South American plant species that are problematic in South Africa. Forty-three insect species...
were recorded on *P. aculeata* during these field trips (I.D. Paterson, unpubl.). Six of these species are discussed below based on the fact that they were either commonly encountered or may hold some promise as potential biological control agents. Specimens are housed at the South African National Collection, Agricultural Research Council-Plant Protection Research Institute (ARC-PPRI), Pretoria, and referred to by Rhodes University (RH) or National Collection (Ac) accession numbers. A leaf-tying moth, *Laxomorpha cambogialis* (Guenée) (formerly under *Epipagis* (Lepidoptera: Pyralidae), which was rejected due to its broad host range, is not included because it was dealt with in a previous review (Klein 1999).

*Maracayia chlorisalis* (Walker) (Lepidoptera: Crambidae) (RH745, RH746, RH774) is considered a promising agent due to the mode of damage and the effect this has on plants in its country of origin. Eggs are deposited on young shoots and the larvae bore into and mine the cortex, pushing excreta through holes to the outside (Klein 1999). The damage sometimes results in shoot death and induces the development of secondary shoots from below the mine. Preliminary host-specificity testing and the known host range of *M. chlorisalis* suggest that it will be restricted to a few genera in the Cactaceae and the moth is therefore likely to be safe for release in South Africa.

However, *M. chlorisalis* is considered a pest on *Hylocereus undatus* Britton & Rose in the State of Puebla, Mexico (CESAVEP 2006) and was also recorded on *Hylocereus ocamponis* (Salm-Dyck) Britton & Rose in Columbia (Zenner de Polania 1990). The fruits of *Hylocereus*, commonly referred to as pitahaya or dragon fruit, are widely consumed in Asia and in some South and Central American countries, and are rapidly becoming popular in the European market (Nerd et al. 2002; Le Bellec et al. 2006). It is traditionally grown in South-East Asia and Central America, and is now considered a promising new crop species elsewhere. It is cultivated extensively in Israel and on a smaller scale in Australia and on Réunion Island (Van Wyk 2005; Le Bellec et al. 2006). Pitahaya is a new crop species grown on a small scale in South Africa (J. Husselman, Institute for Tropical and Subtropical Crops, ARC, pers. comm.). Thus, the introduction of *M. chlorisalis* could cause a conflict of interest between biological control practitioners and pitahaya farmers. Although the pitahaya industry in South Africa is very small at present there is a great deal of potential for pitahaya production in South Africa and neighbouring countries. The possible negative impact of *M. chlorisalis* on South African pitahaya production should not be ignored, but, currently, the benefits of control of *P. aculeata* outweigh the benefits of pitahaya production. *Maracayia chlorisalis* should therefore be considered for release provided host-specificity tests confirm that the insect’s host range is restricted to the Cactaceae.

An undescribed moth species, *cf. Porphyrosela* sp. (Lepidoptera: Gracillariidae: Lithocolletinae) (RH747, RH748), is another potential agent for use in South Africa. It mines the leaves of *P. aculeata*, feeding between the epidermal layers of the leaf blade and pupating within the mine. Members of the Lithocolletinae are usually host specific or live on a suite of closely related plants (Holloway et al. 1987) but observations in the native range suggest that this species feeds on *Talinum paniculatum* Gaertner (Portulacaceae) and *Anredera cordifolia* (Ten.) Steenis (Basellaceae). Leaf-mining species are not usually considered effective biological control agents (Harris 1973; Goeden 1983) and it is possible that damage to the leaves of the plant without destroying the growing tips would have little negative effect on overall plant growth. Although observations in the native range suggest that this species may have a broad host range and inflict little damage, further research is warranted.

If *cf. Porphyrosela* sp. proves to be suitably host specific for release in South Africa it could contribute to control in conjunction with other agents.

An unidentified species of Tenthradinidae (Hymenoptera) was found at coastal sites in Santa Catarina Province, Brazil (26°46′00″S 48°38′27″W; 26°47′26″S 48°35′16″W; 26°55′07″S 48°38′33″W). The larvae are large, solitary, external leaf feeders and the final-instar larva migrates to the base of the plant to pupate in the soil. Although this species has only been found in one localized area, the climatic conditions there were found to match those where *P. aculeata* is invasive in South America. This species should be identified, if possible, prior to host-specificity testing to reveal any available host-range records, and because similar-looking larvae have been observed feeding on *Talinum paniculatum* Gaertner (Portulacaceae) in a nearby region in South America (Misiones, Argentina).

A leaf-galling wasp, *Bruchophagus* sp. (Hymenoptera: Eurytomidae) (AcSN 2784, RH 749, RH 750), was found in Rio de Janeiro and Santa Catarina provinces, Brazil, and in Misiones Province, Argen-
Each wasp larva forms a button gall up to about 5 mm in diameter causing a raised area on the surface of the leaf. A yellow necrosis is sometimes associated with the galls, presumably as the galls age. Even at sites where Bruchophagus sp. was found to be reasonably common, such as north of Rio de Janeiro (22°56'29"S 42°39'57"W) and near Itajai in Santa Catarina Province (26°55'07"S 48°38'32"W), the plants appeared to be unaffected by the galls. The majority of the galls seen were green, so photosynthesis does not appear to be immediately disrupted by the galling action and Bruchophagus sp. by itself is unlikely to be adequately damaging for biological control.

Weevil larvae (Coleoptera: Curculionidae) were found mining the stems of P. aculeata at two coastal sites in Santa Catarina Province, Brazil (26°45'43"S 48°38'49"W; 26°47'19"S 48°35'14"W). The larvae have not been reared to the adult stage to allow identification and nothing is known about the potential host range of the species. It is possible that more than one species of curculionid was responsible for the damage. Because of the mode of damage and the past success of Curculionidae as biological control agents, identification of the species should be considered a priority.

Galled fruits were found at four sites near Foz do Iguaçu in Misiones Province, Argentina (25°41'29"S 54°26'13"W; 25°35'18"S 54°31'08"W; 25°36'24"S 54°29'39"W; 25°40'33"S 54°26'47"W). Affected fruits were enlarged and did not ripen to the orange colour of an undamaged fruit, remaining green and developing a woody exterior and apparently remaining on the plant for an extended period. Hymenopterous larvae found in the fruits, at the first of the four listed sites, were tentatively assumed to be responsible for the galling. The galled fruits did not contain seeds. Seeds of P. aculeata are viable in South Africa (Campbell 1988) and are spread by birds and bats that consume the fruits, enhancing the need for a seed-feeding agent such as this. Maintaining a culture of this species in quarantine may be difficult as a supply of young, actively growing fruits will most likely be needed for oviposition.

Future research into the biological control of P. aculeata

The most promising of the potential agents, based on damage done to the plant in the region of origin, are M. chlorisalis, the tenthredinid and the curculionid stem borers. These species should be considered priority species and collected in southern Brazil, preferably in Rio de Janeiro Province, for further study.

Further intensive general surveys for new potential agents are also needed, especially in areas that have not yet been visited. If no suitable natural enemies are found in the southern regions of the plant’s native distribution, then surveys should be conducted in the northern region of the native distribution where plants are less closely aligned with the South African populations (Paterson et al. 2009). Natural enemies found in the northern region of the native distribution could possibly form new host-plant associations and perhaps, therefore, according to the theory of Hokkanen & Pimentel (1984, 1989), might be more damaging than agents that are taxonomically closer to the South African P. aculeata populations.

Extensive surveys for M. chlorisalis on wild and cultivated pitahaya species and other species of Cactoideae in southern and eastern Brazil are necessary to determine its natural host range and to determine whether M. chlorisalis is likely to be a threat to South African pitahaya farmers. It is possible that there are different forms of M. chlorisalis and that the form present in southeastern Brazil differs from the form which is reported as a crop pest in Central America.

The recent observations of extensive damage by P. guerini at the Weewe Dam site in KwaZulu-Natal, suggests that this species may still have a role to play as a biological control agent, and that releases of P. guerini should continue and post-release evaluations should commence.

**OPUNTIA STRICTA**

Two previous reviews of the biological control of O. stricta in South Africa (Moran & Zimmermann 1991b; Hoffmann et al. 1999) described the formative stages of a programme that has since become highly successful. The two agents deployed, the cactus moth C. cactorum and, especially, the cochineal insect Dactylopius opuntiae (Cockerell) (Hemiptera: Dactylopiidae), continue to suppress O. stricta populations. Opuntia stricta is currently considered under substantial control (Klein 2011).

An ongoing monitoring programme at research sites in the Kruger National Park (25°01'S 31°35'E) has shown that the biomass of O. stricta (measured as cladodes/m²) declined by approximately 90% within six years of the ‘stricta’ biotype of D. opun-
...tics being released and that the cactus has remained at low levels ever since (Fig. 3). The period between release of cochineal insects and the abrupt decrease in biomass of the cactus could have been much shorter were it not for a period of high rainfall and flooding in the region during the late 1990s. Heavy rain destroys cochineal insects by dislodging the immobile females and nymphs from the host plant (Moran & Hoffmann 1987; Moran et al. 1987), causing their populations to dwindle, and allows recovery of the host plants, which flourish while ample moisture is available and damaging insects are scarce. This pattern was apparent between 1998 and 2000 (Fig. 3) when, after an initial escalation in cochineal insect populations and signs of a drop in cactus biomass in 1997, the relative abundance of the cochineal insects declined and the biomass of cactus increased. A period of low rainfall during the early part of the 21st century once again favoured the cochineal and a rapid increase in abundance of the insects was accompanied by a sharp decline in the cactus (Fig. 3). Since 2002 the biomass of cactus has remained at low levels and insecticidal-exclusion experiments have confirmed that both C. cactorum and D. opuntiae are responsible for keeping the weed in check (J.H. Hoffmann et al., unpubl.).

Besides reducing the standing biomass of O. stricta, the insects have also brought about a substantial decline in the mean size of the O. stricta plants in the thickets, from c. 11 cladodes per plant before 2003 to c. 2 cladodes per plant since then (J.H. Hoffmann et al., unpubl.). This has happened because nearly all plants of fruit-bearing size (i.e. >27 cladodes) (Hoffmann et al. 1998a,b; Lotter & Hoffmann 1998) have been destroyed and practically no fruits have been produced in the research plots since 2005 (J.H. Hoffmann et al., unpubl.). The lack of fruits has put a stop to the long range dispersal potential of O. stricta, thereby curbing expansion of the weed into surrounding unaffected areas.

The success of the biological control programme against O. stricta has brought about a substantial decrease in the use of herbicides that otherwise would have had to be used to contain the problem. Limitations on the natural dispersal of D. opuntiae (Foxcroft & Hoffmann 2000) have been overcome by instituting a manual distribution campaign in Kruger National Park. The team of labourers that was previously employed to apply herbicides is now responsible for rearing D. opuntiae in a specially-constructed facility from where consignments of the insects are deployed, both within the park and on surrounding properties (Foxcroft & Hoffmann 2000). The whole management programme against O. stricta in Kruger National Park has focused on maximizing the use of biological control and is now almost entirely reliant on C. cactorum and D. opuntiae (Lotter & Hoffmann 1998; Foxcroft et al. 2004, 2007a,b; Foxcroft & Freitag-Ronaldson 2007; Foxcroft & Rejmanek 2007).

**Biotypes of cochineal insects and hybridization**

The introduction of a distinct biotype of D. opuntiae onto O. stricta in South Africa has opened...
opportunities for there to be cross breeding between this and the already-long-established but different biotype on *O. ficus-indica*. In using the two biotypes, consideration needs to be given as to what might happen when the host plants occur sympatrically and the insects are free to interbreed. Laboratory studies showed that the two biotypes interbreed freely and produce viable progeny that develop equally well on either of their parental host plant species (Hoffmann et al. 2002).

The production of a more-generalist lineage of cochineal could be advantageous for biological control because the insects would have a larger pool of plants on which to develop, enhancing chances of successful host-plant location by the passively-dispersed crawlers and boosting the overall populations of *D. opuntiae*. This potential advantage is negated because the F² and subsequent generations produce progeny which revert to being host specific and at least half of the crawlers that are produced occur on plants on which they cannot develop (Hoffmann et al. 2002; Hoffmann 2004).

Hybridization has happened under field conditions with an outcome that was not anticipated and, under some circumstances, with possible negative consequences for the use of *D. opuntiae* as a biological control agent (Hoffmann 2004). In at least one area where the ‘stricta’ biotype of *D. opuntiae* was introduced onto *O. stricta* plants interspersed with *O. ficus-indica*, ‘ficus’ biotype males have crossed with ‘stricta’ biotype females but not the converse (Hoffmann 2004). Such crossings result in a disproportionate number of crawlers with ‘ficus’ traits hatching on *O. stricta* and not surviving because they are on an unsuitable host. While the consequences of these asymmetrical crosses have not yet been investigated, it is envisaged that the performance of *D. opuntiae* as a biological control agent should not be affected on *O. ficus-indica* but could be curtailed, or even lost altogether, on *O. stricta* populations that are in close proximity to *O. ficus-indica* (Hoffmann 2004). Every effort is currently being made to keep the two biotypes apart and to use only one biotype where *O. ficus-indica* and *O. stricta* occur together.

**CYLINDROPUNTIA FULGIDA VAR. FULGIDA**

*Cylindropuntia fulgida* (Engelm.) F.M.Knuth var. *fulgida* is an upright, much-branched cactus (Fig. 4), the terminal segments of which are easily dislodged. Its cylindrical stem segments are covered in dense clusters of elongate yellow spines, each enclosed in a detachable sheath (Anderson 2001). The flowers are pink to magenta with recurved petals. Fruits remain attached to the plant and new flowers develop from the areoles of the fruit, giving rise to chains of fruit (Benson 1982). The species occurs naturally in the Sonoran Desert of Arizona, and Sonora, Sinaloa and northern Baja California, Mexico. It is commonly known as the chain-fruit cholla or jumping cholla (Anderson 2001) – the latter refers to the ease with which its terminal segments are able to attach themselves to animals or humans that brush past the plant.

Chain-fruit cholla was imported into South Africa, probably as an ornamental, at least as early as the 1940s (De Beer 1986). It is now encroaching upon natural pastoral land and conservation areas in the warm, arid parts of the country, reducing the value and productivity of infested areas (Moran & Zimmermann 1991b) and causing harm, even death, to domestic animals, small antelope and other small mammals and birds. Infestations are localized around Douglas in the Northern Cape Province (Moran & Zimmermann 1991b), as well as on both the South African and Zimbabwean sides of the Limpopo River near Musina in the Limpopo Province (Table 1). Infestations have also been reported from the Western Cape.
North West and Free State provinces of South Africa (Henderson & Zimmermann 2003) (Fig. 5), although some of these records might mistakenly be of the closely related \textit{Cyl. tunicata} (Lehmann) F.M.Knuth or \textit{Cyl. rosea} (DC.) Backeb.

Herbicidal control subsidized by the Department of Agriculture, first using picloram and later MSMA, has been in practice since the late 1970s without much success. The costs involved are prohibitive, particularly for the low-value land on which many of the infestations occur. The Working for Water Programme of the Department of Water Affairs is currently employing people for mechanical and chemical management of the infestations as part of a public works programme. Large parts of the infestation east of Musina have been controlled through this initiative (H. Klein, unpubl.).

**Fig. 4. Cylindropuntia fulgida.** (Drawn by W. Roux; first published in Henderson (1995), and modified in Henderson & Zimmermann (2003), ARC-Plant Protection Research Institute, Pretoria.)

Taxonomy of \textit{Cylindropuntia fulgida} var. \textit{fulgida} and of its associated cochineal insects

In common with many other cactus biological control projects, taxonomic confusion has impeded exploration for and the success of natural enemies used against \textit{Cyl. fulgida} (Moran & Zimmermann 1984), which for many years was incorrectly identified in South Africa as \textit{Cyl. rosea} (= \textit{Cyl. pallida} (Rose) Knuth) (Wells et al. 1986), and earlier as \textit{Cyl. tunicata} (M.J. Wells, unpubl.). Because of previous successes in South Africa and elsewhere, biological control using cochineal insects, \textit{Dactylopius} species, was also the preferred control option against chain-fruit cholla (Moran 1981; Moran & Zimmermann 1984, 1991a; Hosking \textit{et al.} 1988; Zimmermann & Moran 1991; Hoffmann \textit{et al.} 1999). \textit{Dactylopius tomentosus} (Lamark), which was known to attack \textit{Cylindro-
Punica species (Mann 1969), was present in South Africa, having been introduced during 1969 via Australia, and released in 1970 for the successful control of *Cyl. imbricata* (Haw.) F.M.Knuth (Moran & Zimmermann 1991b). Attempts were made to use these populations of *D. tomentosus* (i.e. those of the identical provenance, originating from Australia, and already present in South Africa) against *Cyl. fulgida* var. *fulgida* near Douglas in the Northern Cape during the 1970s. The insects survived and even killed some small plants, but they were ineffective against the large plants (Moran & Zimmermann 1991b) and the infestations of the weed persisted and increased.

During 2000, attempts were made to obtain other provenances of the agent and a consignment of *D. tomentosus* was collected in Mexico from *Cyl. rosea*, which was the species presumed to be problematic in South Africa at the time. This release also did not succeed, creating some doubts about the correctness of the identity of the weed (Mathenge et al. 2009b). Consultation and exchange of material with cactus taxonomists from Mexico and the U.S.A. culminated in the conclusion that the problem cactus in South Africa was indeed *Cyl. fulgida* var. *fulgida*. The initial taxonomic confusion seems to have arisen because some of the plants around Douglas were uncharacteristically small and lacked the chain-fruits, which are so characteristic of *Cyl. fulgida* var. *fulgida*. However, the discovery of plants bearing chains of fruits and floral structures characteristic of *Cyl. fulgida* var. *fulgida* on the Zimbabwean side of the Limpopo River gave the vital clue that led to resolution of the dilemma: in Zimbabwe the cactus was not subject to any form of control and the plants there were reaching their full size and state of maturity, bearing chain fruits, unlike the populations of plants near Douglas where chemical control operations were continually destroying the largest plants in the population and where no chain-fruits were being produced. Furthermore, the subsequent discovery of old photographs from the Douglas area, prior to the start of the chemical control campaign, revealed that large plants with typical chains of fruits had occurred in

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**Fig. 5.** Distribution of *Cylindropuntia fulgida* in South Africa. □, *Cyl. f. var fulgida*; ●, *Cyl. f. var mamillata*. (Drawn by L. Henderson; data source: SAPIA database, ARC-Plant Protection Research Institute, Pretoria.)
the area, confirming the identity of the problem cactus as Cyl. fulgida var. fulgida (Henderson & Zimmermann 2003).

Once the identity of the target weed had been definitively resolved, further shipments of D. tomentosus were obtained from Mexico during 2001. These collections purposefully included populations from several Cylindropuntia species, based on the recent discovery that the cochineal insect, D. opuntiae, comprised at least two biotypes (Volchansky et al. 1999; Githure et al. 1999), implying that D. tomentosus might also have distinct biotypes on the range of host plants with which it is associated. Cochineal insects were collected from Cyl. fulgida var. fulgida between Sonoyta and Puerto Penasco (31°12’N 112°27.5’W), from Cyl. fulgida var. mamillata (Schott ex Engelm.) Backeb. at Hermosillo, in the Sonoran Desert (29°3’N 110°58’W), and, fortuitously, also from the morphologically similar Cyl. cholla (F.A.C. Weber) F.M. Knuth (chain-link cholla) collected near Loreto (23°44’N 110°06’W), and at the CIBNOR research station near La Paz (24°10’10”N 110°17’43”W), both in Baja California Sur.

In South Africa, the biology and host specificity of these provenances of D. tomentosus were investigated (Mathenge et al. 2009a,b) revealing that D. tomentosus had some unique characteristics, e.g. due to a prolonged egg stage, the life cycle of the females is of 63 days duration which is about 17 days longer than that of its congeners. Host-specificity trials confirmed that the different provenances of D. tomentosus behaved as distinct biotypes (Mathenge et al. 2009b; 2010b). Surprisingly, development of the ‘cholla’ biotype (the population of D. tomentosus collected from Cyl. cholla in Baja California Sur) was faster than that of any of the other populations collected from Cyl. fulgida var. fulgida, their crawlers settled and survived more successfully, reached maturity more rapidly, and the resultant females were larger and produced more progeny (Mathenge et al. 2009b). Thus the ‘cholla’ biotype became the agent of choice for release against Cyl. fulgida var. fulgida in South Africa (Mathenge et al. 2009b).

Laboratory hybridization trials between the ‘cholla’ biotype and the ‘imbricata’ biotype of D. tomentosus revealed that the two biotypes were still reproductively compatible, and that hybrid offspring were less host specific than either of the parental lines (Mathenge et al. 2010a), which could be beneficial to biological control. However, since the effects of hybridization on later generations and the influences of the environment on the insects were not tested, it was recommended that the new ‘cholla’ biotype only be released on Cyl. fulgida var. fulgida in areas that are free from cochineal-infested Cyl. imbricata plants, pending further studies in the field.

Investigations using molecular techniques demonstrated that the D. tomentosus populations collected from Cyl. fulgida var. fulgida, Cyl. fulgida var. mamillata and Cyl. imbricata had closer phylogenetic relationships with each other than with populations from Cyl. cholla (Mathenge 2010; Mathenge et al. in prep.). The largest genetic divergence was recorded in populations of D. tomentosus from Cyl. rosea suggesting that the ‘rosea’ biotype may justify a different taxonomic status, such as a host race or sibling species, and explains why this provenance had failed earlier on in the programme.

Impact of the cochineal insects on Cyl. fulgida var. fulgida

The ‘cholla’ biotype of D. tomentosus was released on a large infestation of Cyl. fulgida var. fulgida on the farm Kleinplaas near Douglas, Northern Province (Table 1) but was less damaging to the weed than had been expected. Suspicions that the insect culture might have inadvertently been contaminated with the biotype from Cyl. imbricata during mass-rearing prompted acquisition of new shipments of the insects for release. During September 2007 two populations of D. tomentosus were collected from Cyl. cholla growing at the two Baja California Sur sites in Mexico. In South Africa the colony was cleared of predators, parasitoids and contaminants under quarantine conditions at Rietondale, Pretoria, and mass-reared for release during 2008 at the localities listed in Table 1.

Four months after the release near Musina, all of the inoculated plants were heavily colonized with cochineal insects, cladodes were falling from the plants, small plants were dying and the insects had dispersed up to 5 m from the release sites. By November 2009, a year after release, many plants were dead, and it was difficult to find plants in the area that were not heavily colonized by cochineal insects. By June 2010, the cochineal insects had dispersed throughout the 87-ha infestation, the majority of small plants were dead, and only a few large cactus plants with woody stems still survived,
although most of their cladodes had detached and died. Heavy summer rains at the time washed the cochineal insects from the exposed sides of the surviving plants but a substantial portion of the cochineal population survived on the underside and protected parts of the stems and cladodes.

As a result of the unexpectedly rapid proliferation of the cochineal insects at Musina, the opportunity was missed to monitor the earliest stages in the establishment, including population growth and dispersal of the insect, and to measure the early effects on the target weed. Long-term monitoring of the fate of later releases in the Northern Cape, where the cactus is evenly distributed, is planned.

A visit to Douglas in the Northern Cape during July 2010 showed that the damage from cochineal insects was less marked than it had been at Musina. There was considerable new growth on many of the large, woody-stemmed cactus plants but practically all the small plants and detached segments were dead or dying. The new growth on all monitored plants was already infested with cochineal insects (H. Klein, unpubl.). The superior performance of the cochineal insects at Musina compared to Douglas is probably related to Musina’s higher winter temperatures and lower rainfall, both of which are known to favour cochineal insect development (Zimmermann & Moran 1982; Moran & Hoffmann 1987; Moran et al. 1987).

During March 2009, the Working for Water clearing team at Musina was advised about the Cyl. fulgida var. fulgida infestations and they agreed to harvest cactus cladodes colonized by the cochineal insects and to redistribute them, instead of applying herbicides. Cylindropuntia fulgida var. fulgida is now considered under complete control (Klein 2011).

CEREAUS JAMACARU

The initiation of biological control against the columnar cactus, Cereus jamacaru, commonly known as queen of the night cactus (Fig. 6), was reviewed by Moran and Zimmermann (1991b) and by Klein (1999). This project is the secondary consequence of the release of two agents, the mealybug, Hypogeococcus pungens Granara de Willink (Hemiptera: Pseudococcidae), referred to in earlier literature as H. festerianus (Lizer & Trelles), and the stem borer Nealcidion cereicola (Fisher) (formerly under Alcidion) (Coleoptera: Cerambycidae), against another cactus, Harrisia martinii in the same subfamily, Cactoideae. As expected, both of these agents readily accepted Ce. jamacaru as a host.

Nealcidion cereicola has become established at only a few sites, but is very damaging where populations reach high levels. By weakening the structural tissues of the cactus, they cause large stem sections to break off, or the entire plant to collapse.

By contrast, H. pungens has been actively redistributed throughout the range of Ce. jamacaru in South Africa (Fig. 7). In most parts of the country, this agent keeps Ce. jamacaru under effective control, and all other control measures have been discontinued. When the mealybug becomes established on a cactus plant, the insects congregate in any area of the plant which is actively growing, where they cause abnormal growth, especially on the stem tips, which become swollen and spiny. These lumps apparently result from the proliferation of thin, twisted shoots with shortened internodes and closely-spaced spines. Each infested plant dies eventually: young plants succumb before having reached flowering age, whereas plants that only become infested when they are several metres tall, may take up to four years to die. However, even on such large plants, fruit production is markedly reduced almost from the time that the insects become established, since the mealybugs aggregate on the developing flowers. Generally, one large specimen of C. jamacaru, infested with H. pungens, is sufficient to ensure that all seedlings germinating in the area become infested and die (H. Klein, pers. obs.). No formal quantitative post-release evaluations have been undertaken on this biological control project but the weed is considered under complete control (Klein 2011).

In KwaZulu-Natal Province, the cactus has been reported to tolerate H. pungens damage better than in the rest of the country (R. Brudvig, pers. comm.). It is not clear whether this is a result of the subtropical climate or a combination of other factors, including parasitism and predation. Predation of H. pungens by coccinellid beetles such as Chilocorus species and parasitism by the encyrid wasp, Homalotylus flaminius (Dalman) (Hymenoptera: Encyrtidae), have been observed in South Africa (Danninger 2002), but their impact has not been quantified. Cereus jamacaru plants growing in KwaZulu-Natal might differ taxonomically from those in other parts of the country;
for example, it is suspected that *Ce. hildmannianus* K. Schum. (= *Ce. uruguayanus* Ritter ex Kiesling) might be present in South Africa. However, since *H. pungens* (as *H. festerianus*) is known to feed widely within the subfamily Cactoideae (McFadyen & Tomley 1981), this is unlikely to be a limiting factor in the biological control of *C. jamacaru*.

**CONCLUSIONS**

In retrospect, after a century of biological control projects against cactus weeds in South Africa – from the successful control of *O. monacantha* in 1913 (Lounsbury 1915) until the equally successful control of *Cyl. fulgida var. fulgida* in 2010, it is possible to identify three distinct stages in cactus biological control which in aggregate have resulted in relatively high levels of success when compared to biological control projects against other plant families in South Africa.

The earliest cactus biological control projects involved no research by South Africans. Instead, the first project exploited the inadvertent control of a cactus by a mistakenly introduced cochineal insect species to India in 1795 (Zimmermann et al. 2009), and several subsequent projects benefited from this precedent in India, and from subsequent research and implementation of biological control carried out in Australia. The second stage, however, did include several South African initiatives, beginning with the introduction of a weevil, *Microstigmus spinolae* (Gyllenhal) (Coleoptera: Curculionidae), for the biological control of *O. ficus-indica* in 1948 (Annecke & Moran 1978).

Contrary to the prediction by Moran & Zimmermann (1985) that ‘there seems to be little chance of

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**Fig. 6. Cereus jamacaru.** (Drawn by G. Condy; first published in Henderson (1995), ARC-Plant Protection Research Institute, Pretoria.)
finding new dactylopiid species in the Americas, [and that] the emphasis in the biological control of cacti should shift to the use of pathogens in combination with plant-sucking bugs’, the third stage, spanning the period from 1999 to 2010, involved almost exclusively the use of new cochineal insect biotypes of two *Dactylopius* species that provided excellent control of two cactus weeds, *O. stricta* and *Cyl. fulgida var. fulgida*, respectively. With the exception of *P. aculeata*, the future of cactus biological control in South Africa is likely to rely further on host-adapted cochineal biotypes for the control of incipient cactus weeds such as *Opuntia humifusa* (Rafinesque) Rafinesque, *Cyl. rosea*, *Cyl. tunicata* and *Opuntia engelmannii* Salm-Dyck ex Engelm. Another emerging cactus weed in South Africa, *O. salmiana* J.Parm ex Pfeiff., could possibly be controlled using its own monophagous cochineal insect, *Dactylopius salmianus* De Lotto (De Lotto 1974).

The relatively depauperate fauna of phytopagous insects associated with *P. aculeata* contrasts with the 150–160 phytophagous insect species associated with the more than 175 succulent cacti surveyed so far (Mann 1969; Zimmermann et al. 1979). The paucity of species on *P. aculeata* is unexpected given that the host plants are woody scrambling shrubs that have true leaves and seemingly offer many niches and an abundant resource for leaf-feeding and stem- or twig-boring insects. The situation calls for further exploration and study.

The taxonomic complexities and inconsistencies within in the family Cactaceae (Hunt 2006) have been a long-standing source of confusion for entomologists who study cactophagous insect species associated with very specific cactus taxa. For example *Acanthocereus tetragonus* (L.) Humik. (Cactaceae), which is naturalized in South Africa, and has the tendency to be invasive, is recorded as one of six species in the genus by Anderson (2001), while Hunt (2006) lists only one species in the genus *Acanthocereus*. There are several cases where the biological control of problematic cactus species has been impeded by an inadequate understanding of the taxonomy of the group under investigation (Moran et al. 1976). The work of the International
Cactaceae Systematics Group of the International Organization for Succulent Plant Studies, and the availability of modern molecular taxonomic techniques, is making substantial contributions towards a new practical classification of the Cactaceae and is resulting in improved nomenclatural consistency and reliability. This will be advantageous to future biological control projects against cactus weeds, in particular.

Extensive infestations of dominantly *Opuntia* species are becoming increasingly problematic in other African countries. These include *O. stricta* var. *stricta* and var. *dillenii* in Madagascar, Namibia, Kenya, Ethiopia, Yemen and Saudi Arabia, and *O. elatior* Mill. Gard. in Kenya. There is no doubt that the South African experiences with host-specific cochineal insect biotypes will be the foundation of successful biological control in these countries, should they opt for biological control. Recent experiences in research and implementation of biological control could be of particular and immediate benefit to South Africa’s immediate neighbours, Zimbabwe and Namibia, which share many of the same problems of invasive cactus species and where many of the biological control agents that have been used could be effective.

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